exo,endo and exo,exo isomers of 6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> and its halogenated derivatives. Molecular structures of exo,endo- and exo,exo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> and exo-6,endo-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-arachno-B<sub>10</sub>H<sub>11</sub>

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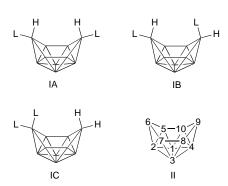
Reaction of PMe<sub>2</sub>Ph with *nido*-B<sub>10</sub>H<sub>14</sub> at *ca.* 200 K gave *exo*,*exo* and *exo*,*endo* isomers of 6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-*arachno*-B<sub>10</sub>H<sub>12</sub> which were separated chromatographically. Both forms were confirmed crystallographically. The compounds 2-Br-*nido*-B<sub>10</sub>H<sub>13</sub> and 2,4-Cl<sub>2</sub>-*nido*-B<sub>10</sub>H<sub>12</sub> gave exclusively *exo*,*endo* forms at *ca.* 200 K with the 2-Br product consisting of equal amounts of 6-*endo*-9-*exo* and 6-*exo*-9-*endo*-(PMe<sub>2</sub>Ph)<sub>2</sub> isomers, demonstrating no directional control by the 2-Br substituent. The relative configurations of the two brominated isomers were confirmed by a crystallographic study of the 6-*exo*-9-*endo* species. Both the monobrominated *exo*,*endo* isomers gave the *exo*,*exo* counterpart on heating, but the 2,4-Cl<sub>2</sub> species is more robust, with apparent decomposition on extended heating, and no evidence for the formation of *exo*,*exo*-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>B<sub>10</sub>H<sub>10</sub>.

Since the discovery in 1957 that *nido*-decaborane,  $B_{10}H_{14}$ , readily reacts with nucleophilic bases L to give *arachno* species 6,9- $L_2B_{10}H_{12}$  1 [equation (1)]<sup>1</sup> the reaction has become very well

$$B_{10}H_{14} + 2L \longrightarrow L_2B_{10}H_{12} + H_2$$
 (1)

recognized,2 not least because of the importance of these species (where L is a weaker base) as intermediates in closo twelve-vertex dicarbaborane formation.<sup>3</sup> The original crystallographically characterized examples of these L<sub>2</sub>B<sub>10</sub>H<sub>12</sub> species 1 were for L = MeCN and  $SMe_2$ , and these exhibited an exo, exobis(ligand) disposition IA.4,5 This configuration is also shown by more recent crystallographic studies, 6,7 and it seems reasonable to presume that this is the most stable thermodynamic configuration. However, in 1987 it was reported<sup>8</sup> that NMR spectroscopy showed that careful addition of the ligand PMe<sub>2</sub>Ph to nido-B<sub>10</sub>H<sub>14</sub> at lower temperatures gave substantial quantities of the exo,endo 6,9 isomer 1b of configuration IB as well as the expected exo,exo isomer 1a. This was not associated with any crystallographic work. The exo,endo isomer 1b isomerized to the exo,exo 6,9 isomer 1a of configuration IA on mild heating. More recently it was reported that, where L = pyridine(py), a third type of isomer can be formed, the exo,endo 6,6 isomer of configuration IC.9 This also has not yet been substantiated crystallographically, the only  $(py)_2B_{10}H_{12}$  isomer so investigated being the conventional exo,exo 6,9 isomer of configuration IA.7 We have now investigated our PMe<sub>2</sub>Ph system in more detail, with the intention of crystallographic confirmation of the exo,endo 6,9 isomeric form IB, and, with a view to the examination for any exolendo directional control or other isomer preferences, using the halogenated nido-decaboranes  $2-BrB_{10}H_{13}$  and  $2,4-Cl_2B_{10}H_{12}$  as starting substances. We now report results from these investigations. The numbering system used for the nido and arachno ten-vertex geometry is in schematic II.

# **Results and Discussion**

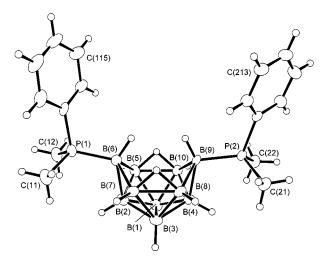


respectively. At the time the formation of the exo,endo isomeric forms of this type of arachno species 6,9-L2B10H12 had not previously been noted. The exo, endo isomer 1b was readily identified in this first instance by NMR spectroscopy on the exo,exol exo,endo mixture of compounds 1a and 1b.8 We have now found in this work that chromatography on silica gel G using hexane-CH<sub>2</sub>Cl<sub>2</sub> (1:9) as solvent can result in the isolation of the two isomers in pure form, isolatable in yields of 57 and 34% respectively. Crystallization from CH<sub>2</sub>Cl<sub>2</sub>-hexane at room temperature gave single crystals of each that were suitable for X-ray diffraction analysis. The crystallographically determined molecular structure of the exo,exo isomer 1a (Fig. 1 and Tables 1 and 2)<sup>10</sup> confirms this isomer to have the two phosphine ligands in exo,exo dispositions (schematic IA), whereas the structure of 1b (Fig. 2 and Tables 1 and 2) clearly confirms its formulation as the exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> isomer (schematic IB) and obviously precludes, for example, the 6,6-exo,endo configuration IC.

For the *exo*,*exo* isomer 1a there are two crystallographically independent molecules (designated A and B) in the asymmetric fraction of the unit cell. The borane clusters of these two molecules are essentially identical within experimental error; they differ in the conformations of the organic moieties bound to phosphorus. Most significant are the twists of the phosphorus-bound phenyl rings about the P–C (Ph) vectors. The ring bound to P(1) in molecule B is twisted by 66.8(4)° relative to that in A, whilst that bound to P(2) in B is twisted by 88.0(4)° relative to that in A. The two molecules of compound 1a and that of 1b all have the classical *arachno* ten-vertex character typical of such 6,9-bis(ligand)decaborane species, and in particular have

Table 1 Selected interatomic distances (Å) and angles (°) for the two crystallographically independent molecules (A and B) of exo, exo,

	1a (A)	1a (B)	1b	2 (X)	2 (Y)
B(2)-B(5)	1.751(4)	1.754(4)	1.765(2)	1.72(2)	1.75(2)
B(2)-B(6)	1.744(5)	1.740(4)	1.735(2)	1.75(2)	1.77(2)
B(2)-B(7)	1.758(5)	1.752(4)	1.758(2)	1.72(2)	1.71(2)
B(5)-B(6)	1.865(5)	1.867(4)	1.851(2)	1.90(2)	1.96(2)
B(6)-B(7)	1.860(4)	1.877(4)	1.864(2)	1.89(2)	1.86(2)
B(6)-P(1)	1.912(3)	1.925(3)	1.9145(14)	1.888(13)	1.918(13)
B(4)-B(8)	1.748(5)	1.760(4)	1.758(2)	1.71(2)	1.74(2)
B(4)-B(9)	1.748(4)	1.732(4)	1.762(2)	1.74(2)	1.78(2)
B(4)-B(10)	1.766(4)	1.755(5)	1.756(2)	1.75(2)	1.75(2)
B(8)-B(9)	1.876(4)	1.859(4)	1.888(2)	1.89(2)	1.88(2)
B(9)-B(10)	1.873(4)	1.866(4)	1.883(2)	1.84(2)	1.87(2)
B(9)-P(2)	1.925(3)	1.914(3)	1.941(2)	1.890(14)	1.884(12)
B(5)-B(10)	1.869(4)	1.879(4)	1.892(2)	1.89(2)	1.86(2)
B(7)-B(8)	1.875(5)	1.871(4)	1.879(2)	1.90(2)	1.90(2)
B(2)-B(6)-P(1)	110.6(2)	107.0(2)	111.20(8)	112.6(8)	124.5(8)
B(5)-B(6)-P(1)	119.3(2)	117.3(2)	118.23(8)	95.5(6)	95.4(7)
B(7)-B(6)-P(1)	118.6(2)	116.5(2)	120.18(8)	95.0(7)	96.6(7)
B(4)-B(9)-P(2)	105.8(2)	111.1(2)	155.86(10)	124.1(9)	125.4(8)
B(8)-B(9)-P(2)	116.4(2)	117.1(2)	111.33(8)	95.8(8)	96.3(7)
B(10)-B(9)-P(2)	116.8(2)	120.3(2)	110.42(8)	95.4(7)	96.7(7)
B(6)-B(5)-B(10)	113.2(2)	113.5(2)	112.88(9)	105.9(9)	105.5(9)
B(5)-B(6)-B(7)	103.8(2)	104.1(2)	104.52(9)	105.1(8)	103.5(8)
B(6)-B(7)-B(8)	113.1(2)	112.3(2)	113.22(9)	107.4(9)	107.3(8)
B(7)-B(8)-B(9)	113.2(2)	113.4(2)	115.96(9)	106.3(9)	106.0(8)
B(8)-B(9)-B(10)	103.7(2)	104.8(2)	101.91(9)	107.1(9)	106.5(8)
B(9)-B(10)-B(5)	113.3(2)	111.9(2)	116.10(9)	108.3(9)	108.3(8)
B(1)-B(2)-B(6)	115.4(2)	116.4(2)	115.21(10)	114.9(9)	113.8(8)
B(3)-B(2)-B(6)	115.7(2)	116.5(2)	115.74(10)	114.4(9)	112.4(8)
B(1)-B(4)-B(9)	116.3(2)	115.6(2)	118.32(10)	114.2(10)	113.9(8)
B(3)-B(4)-B(9)	116.9(2)	115.9(2)	118.38(10)	113.2(10)	113.0(8)



**Fig. 1** An ORTEP-type <sup>10</sup> drawing of the crystallographically determined molecular structure of one of the crystallographically independent molecules (A) of  $exo_1exo_2$ -6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>- $arachno_2$ -B<sub>10</sub>H<sub>12</sub> **1a** with thermal ellipsoids shown at the 40% probability level. Phenyl rings C(n1i) (n=1 or 2, i=1-6) are numbered cyclically and hydrogen atoms have been assigned a small, arbitrary radius for clarity. Molecule B only differs significantly from A in the orientation of the phosphorus-bound substituents

B(5)–B(10) and B(7)–B(8) connectivities [range 1.869(4)–1.892(2) Å] characteristically shorter than the corresponding distance (typically *ca.* 2.00 Å) in *nido*-decaboranyl clusters. The presence of a hydrogen atom bridging each of these vectors is further consistent with their *arachno* constitution.

The effect upon structure of an exo versus an endo phosphine substituent may be evaluated by comparison of the

**Table 2** Additional interatomic distances (Å) and angles (°) for the two crystallographically independent molecules (A and B) of exo, exo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> **1a** and exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> **1b** 

	1a (molecule A)	1a (molecule B)	1b
B(1)-B(2)	1.769(5)	1.762(4)	1.769(2)
B(1)-B(3)	1.812(5)	1.819(5)	1.816(2)
B(1)-B(4)	1.765(5)	1.769(5)	1.755(2)
B(1)-B(5)	1.766(5)	1.781(5)	1.779(2)
B(1)-B(10)	1.788(4)	1.778(5)	1.791(2)
B(2)-B(3)	1.766(5)	1.769(5)	1.765(2)
B(3)-B(4)	1.767(4)	1.767(5)	1.754(2)
B(3)-B(7)	1.783(5)	1.781(4)	1.774(2)
B(3)-B(8)	1.788(5)	1.783(5)	1.789(2)

Cluster B-B-B acute angles are within the ranges: **1a** (A) 57.3(2)–64.9(2), **1a** (B) 57.0(2)–65.0(2) and **1b** 57.15(7)–64.88(8)°.

selected geometric parameters given in Table 1, in which salient distances and angles for compounds 1a (A and B) and 1b are listed. Also included for comparison are the corresponding values for what formally constitutes an endo,endo analogue, the [endo,endo-6,9-μ-(PPh<sub>2</sub>)-arachno-B<sub>10</sub>H<sub>12</sub>] anion 2 of schematic structure III (which has two crystallographically independent molecules X and Y in its crystallographically examined [PMePh<sub>3</sub>]<sup>+</sup> salt). 11 Unfortunately the reported structure of this endo, endo species 2 is insufficiently accurate for a detailed comparison with the present compounds. It does appear, however, that the boron-phosphorus distances B(6)-P(endo) and B(9)-P(endo) in compound 2 [1.884(12) to 1.918(13) Å] tend to be rather shorter than those in 1a and 1b. This might equally be a consequence of stereochemical constraints arising from the bridge, or of the differing chemical nature of this anion.

Table 3 Measured <sup>11</sup>B and <sup>1</sup>H NMR chemical shift data for  $exo_1$ ,  $exo_2$ ,  $exo_3$ ,  $exo_4$ ,  $exo_5$ ,  $exo_6$ ,  $exo_$ 

	3a b		3b c		3c <sup>d</sup>	
Position	$\delta(^{11}B)$	δ( <sup>1</sup> H)	$\delta(^{11}B)$	δ( <sup>1</sup> H)	$\delta(^{11}B)$	δ( <sup>1</sup> H)
2	+1.0	_	+3.9	_	+9.8	_
4	-4.0	+2.15	+6.7	$+2.95^{e}$	+0.6	+2.25
1,3	-36.5	+1.00	-32.2	+1.28	-32.1	+1.27
6	$-33.6^{f}$	-1.05	$-33.1^{g}$	-1.53	$-27.7^{h}$	+1.65
9	ca. $-36^{i}$	-1.04	$-31.0^{g}$	+0.99	$-35.6^{f}$	-1.50
5,7	-17.9	+2.06	-13.9	+2.16	-13.7	+1.85
8,10	-19.6	+1.63	-19.0	+1.65	-17.6	+1.95

<sup>a</sup> In CDCl<sub>3</sub> at 294 K;  $\delta$ (<sup>1</sup>H) related to directly bound B positions by <sup>1</sup>H-{<sup>11</sup>B(selective)} spectroscopy; any fine structure noted for <sup>1</sup>H resonances is observed under conditions of complete {<sup>11</sup>B(broad-band noise)} decoupling. <sup>b</sup>  $\delta$ (<sup>31</sup>P) ca. -2 (br); additionally  $\delta$ (<sup>1</sup>H)[μ-H(5,10 and 7,8)] -4.16,  $\delta$ (<sup>1</sup>H)(PMe<sub>2</sub>) +1.88 with <sup>2</sup>J(<sup>31</sup>P-<sup>1</sup>H) = 11.5 Hz and  $\delta$ (<sup>1</sup>H)(PPh) +7.50 to +7.73. <sup>c</sup>  $\delta$ (<sup>31</sup>P) ca. +11 and -2 (both broad); additionally  $\delta$ (<sup>1</sup>H)[μ-H(5,10 and 7,8)] -4.96 with <sup>2</sup>J(<sup>31</sup>P-<sup>1</sup>H) ca. 10,  $\delta$ (<sup>1</sup>H)(PMe<sub>2</sub>) +1.77 with <sup>2</sup>J(<sup>31</sup>P-<sup>1</sup>H) = 11.8 and +1.63 with <sup>2</sup>J(<sup>31</sup>P-<sup>1</sup>H) = 13.0 Hz and  $\delta$ (<sup>1</sup>H)(PPh) +7.29 to +7.73. <sup>d</sup>  $\delta$ (<sup>31</sup>P) ca. +9 and 0 (both broad); additionally  $\delta$ (<sup>1</sup>H)[μ-H(5,10 and 7,8)] -4.83 with <sup>2</sup>J(<sup>31</sup>P-<sup>1</sup>H) ca. 14,  $\delta$ (<sup>1</sup>H)(PMe<sub>2</sub>) +1.83 with <sup>2</sup>J(<sup>31</sup>P-<sup>1</sup>H) = 11.5 Hz and  $\delta$ (<sup>1</sup>H)(PPh) +7.30 to +7.73. <sup>e</sup> Coupling <sup>3</sup>J(<sup>31</sup>P-<sup>1</sup>H) ca. 23 Hz. <sup>f</sup> Coupling <sup>1</sup>J(<sup>31</sup>P-<sup>11</sup>B) ca. 130 Hz; not measurable more accurately due to overlap with <sup>11</sup>B(1,3). <sup>h</sup> Coupling <sup>1</sup>J(<sup>31</sup>P-<sup>11</sup>B) ca. 125 Hz. <sup>i</sup> Not measurable more accurately due to overlap with <sup>11</sup>B(1,3).

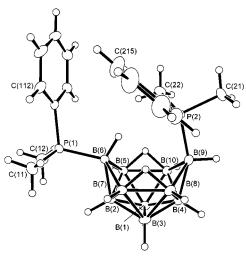
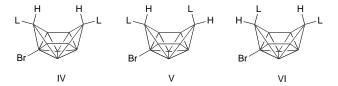


Fig. 2 An ORTEP-type  $^{10}$  drawing of the crystallographically determined molecular structure of *exo,endo-*6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-*arachno*-B<sub>10</sub>H<sub>12</sub> 1b drawn with the same conventions as in Fig. 1



In the exo,endo compound 1b significant structural effects attributable to the endo ligand appear to be localized around its point of attachment, and there appears to be a systematic weakening of the bonding around the endo-substituted boron atom B(9). Thus the boron-phosphorus distance B(9)-P(2)-(endo) in 1b is 1.941(2) Å, significantly longer than the B(6)-P(exo) and B(9)-P(exo) distances in **1a** and **1b** [range 1.912(3)–1.925(3) Å]. The three interboron connectivities adjacent to this B(9)-P(2)(endo) linkage in compound 1b are also elongated relative to the corresponding separations in the 'exo' cluster fragments of 1a and 1b. Specifically, B(4)-B(9) is 1.762(2) Å in **1b** compared to a range of 1.732(4)–1.748(4) Å for the other B(2)–B(6) and B(4)–B(9) distances in **1a** and **1b**, and B(8)-B(9) and B(9)-B(10) are 1.888(2) and 1.883(2) Å respectively in 1b compared to a range of 1.851(2)-1.877(4) Å for the other B(8)-B(9), B(9)-B(10), B(5)-B(6) and B(6)-B(7) distances in 1a and 1b. Moreover, there is a small but significant enlargement [2.5(2) to  $4.2(2)^{\circ}$ ] of the B(7)-B(8)-B(9) and B(9)-B(10)-B(5), angles, and similarly [1.4(2) to 3.2(2)°] of the B(1)–B(4)–B(9) and B(3)–B(4)–B(9) angles, at the endo-



substituent end of compound **1b** relative to the equivalent parameters in the *exo*-configured fragments of **1a** and **1b**. All of these structural effects could be perhaps be attributable to the minimization of steric factors arising from organic groups associated with the *endo*-phosphine substituent in isomer **1b**. However, there must be an underlying electronic contribution to these features, as these deviations appear to be limited strictly to the *endo* end of the cluster in the *exo*,*endo* isomer **1b**. The relative weakness of these linkages to the *endo*-substituted boron atoms are further manifested in the relatively facile thermal isomerization of *endo*,*exo* **1b** to generate the *exo*,*exo* isomer **1a** of greater thermodynamic stability.

Having unequivocally confirmed the exo,endo structural type, it was of interest to examine the reaction of the halogenated derivative 2-Br-nido-B<sub>10</sub>H<sub>13</sub> with PMe<sub>2</sub>Ph under the same conditions for any differential influence that the 2-bromo substituent might have in dictating a 6,9 (structure V) versus a 9,6 (structure V) ver

In the event, addition of PMe<sub>2</sub>Ph to 2-Br-nido-B<sub>10</sub>H<sub>13</sub> at ca. 200 K resulted in clean formation of two products, **3b** and **3c**, in equimolar ratio within experimental error by integrated <sup>11</sup>B NMR spectroscopy. These were isolated by TLC and identified and distinguished by <sup>1</sup>H and <sup>11</sup>B NMR spectroscopy (Table 3) as the 6,9-exo,endo and 9,6-exo,endo isomers of 6,9-(PMe<sub>2</sub>Ph)-2-Br-arachno-B<sub>10</sub>H<sub>11</sub> (compounds **3b** and **3c** respectively). The differential NMR assignment was confirmed by single-crystal X-ray diffraction work on the 6,9-exo,endo isomer **3b**. No other significant products were detectable. In particular the conventionally structured 6,9-exo,exo isomer **3a** (schematic **IV**) was absent, indicating that an effect of the 2-bromo substituent is to divert all of the kinetic product into an exo,endo configuration. This tendency was confirmed by the examination of the

**Table 4** Measured  $^{11}$ B and  $^{1}$ H NMR data for exo, endo-6,9-(PMe $_{2}$ Ph) $_{2}$ -2,4-Cl $_{2}$ -arachno-B $_{10}$ H $_{10}$ 4b $^{a}$ 

Assignment		$\delta(^{11}B)$	$\delta(^{1}H)^{h}$
2 or 4		+15.1	_
2 or 4		+8.8	_
1,3		-30.9	+1.49
6		$-35.4^{c}$	-1.44
9	ca.	$-30^{d}$	+1.45
5,7		-14.4	+2.10
8,10		-19.0	+1.82

<sup>a</sup> In CDCl<sub>3</sub> at 294 K; additionally  $\delta(^{31}P)$  ca. +10 and -1 (both broad);  $\delta(^{1}H)$  related to directly bound B positions by  $^{1}H$ -{ $^{11}B$ (selective)} spectroscopy; any fine structure noted for the  $^{1}H$  resonances is observed under conditions of complete { $^{11}B$ (broad-bond noise)} decoupling. <sup>b</sup> Additionally  $\delta(^{1}H)[\mu$ -H(5,10 and 7,8)] -4.73 with  $^{2}J(^{31}P^{-1}H)$  ca. 12,  $\delta(^{1}H)(PMe_{2})$  +1.70 with  $^{2}J(^{31}P^{-1}H)$  = 11.5 Hz and  $\delta(^{1}H)(PPh)$  +7.21 to +7.70.  $\epsilon^{1}J(^{31}P^{-11}B)$  ca. 125 Hz. <sup>d</sup> Not measurable more accurately due to overlap with  $^{11}B(1,3)$ .

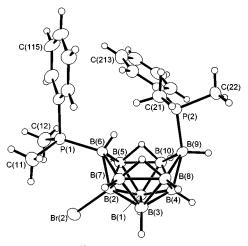
**Table 5** Selected interatomic distances (Å) and angles (°) for exo-6,endo-9- $(PMe_2Ph)_2$ -2-Br-arachno- $B_{10}H_{11}$  3

Br(2)-B(2)	1.986(4)	B(3)–B(7)	1.790(6)
P(1)-B(6)	1.914(4)	B(3)-B(8)	1.796(6)
B(1)-B(4)	1.742(6)	B(4)-B(10)	1.759(6)
B(1)-B(2)	1.754(6)	B(4)-B(9)	1.759(6)
B(1)-B(5)	1.773(6)	B(4)-B(8)	1.759(6)
B(1)-B(10)	1.780(6)	B(5)-B(6)	1.879(6)
B(1)-B(3)	1.822(6)	B(5)-B(10)	1.887(6)
B(2)-B(6)	1.717(6)	B(6)-B(7)	1.846(5)
B(2)-B(5)	1.747(6)	B(7)-B(8)	1.906(6)
B(2)-B(3)	1.755(6)	B(8)-B(9)	1.869(6)
B(2)-B(7)	1.766(5)	B(9)-B(10)	1.882(6)
B(3)-B(4)	1.744(7)		
	` ^		
B(6)-B(2)-B(1)	117.9(3)	B(2)-B(6)-P(1)	118.1(3)
B(6)-B(2)-B(3)	117.0(3)	B(5)-B(6)-P(1)	121.8(2)
B(1)-B(2)-Br(2)	116.5(3)	B(7)-B(6)-P(1)	122.1(3)
B(3)-B(2)-Br(2)	117.2(3)	B(6)-B(7)-B(8)	113.9(3)
B(5)-B(2)-Br(2)	118.8(2)	B(9)-B(8)-B(7)	117.4(3)
B(6)-B(2)-Br(2)	115.5(3)	B(8)-B(9)-B(10)	101.8(3)
B(7)-B(2)-Br(2)	120.0(3)	B(4)-B(9)-P(2)	158.0(3)
B(1)-B(4)-B(9)	118.7(3)	B(8)-B(9)-P(2)	111.1(2)
B(3)-B(4)-B(9)	118.6(3)	B(10)-B(9)-P(2)	112.2(2)
B(6)-B(5)-B(10)	113.8(3)	B(9)-B(10)-B(5)	117.5(3)
B(7)-B(6)-B(5)	104.5(3)		

Cluster B-B-B acute angles are within the range 56.4(2)-64.7(2)°.

same reaction using 2,4-Cl<sub>2</sub>-nido-B<sub>10</sub>H<sub>12</sub> as starting substrate. At ca. 200 K with PMe<sub>2</sub>Ph this gave a quantitative conversion into exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>-arachno-B<sub>10</sub>H<sub>10</sub> (compound **4b**, schematic structure **VII**). Thus there was no evidence for the symmetrical exo,exo isomer **4a**, nor for any significant quantities of {B<sub>10</sub>H<sub>10</sub>Cl<sub>2</sub>} residues other than the 2,4-dichloro configuration. That reaction (1) proceeds without a rearrangement of the {B<sub>10</sub>} skeleton tends therefore to be confirmed. The NMR data for exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>-arachno-B<sub>10</sub>H<sub>10</sub> (**4b**) are in Table 4.

As mentioned in the previous paragraph, the 6-exo,9-endo isomer of the 2-brominated species, compound **3b**, has been examined by a single-crystal X-ray diffraction analysis (Fig. 3 and Table 5), which unequivocally establishes this isomer to be of structure type **V** and confirms our analysis of the NMR spectra of **3b** and, by implication, **3c**. The hydrogen-bridged connectivities B(5)–B(10) and B(7)–B(8) [1.887(6) and 1.906(6)

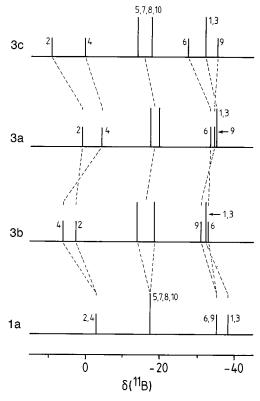


**Fig. 3** An ORTEP-type <sup>10</sup> drawing of the crystallographically determined molecular structure of exo-6,endo-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-arachno-B<sub>10</sub>H<sub>11</sub> **3b** drawn with the same conventions as in Fig. 1

Å respectively] are comparable to those in 1a and 1b and again confirm the arachno character of the cluster. Cluster flexing around the B(9) atom of the kind seen in 1b, which we have associated with the presence of an endo versus an exo phosphine substituent, is also observed in compound 3b. Specifically, (a) the B(9)-P(endo) distance of 1.928(4) Å in 3b is towards the long end of the range established above for B(6)-P(exo) and B(9)-P(exo) distances [1.912(3)–1.925(3) Å], (b) the interboron distance B(4)–B(9) of 1.759(6) Å is much longer than the range of equivalent distances in the exo-substituted fragments of 1a and **1b** [1.732(4)–1.748(4) Å], and (c) there are even greater increases in the B(7)-B(8)-B(9), B(9)-B(10)-B(5), B(1)-B(4)-B(4)B(9) and B(3)-B(4)-B(9) angles in 3b compared to the equivalent values for the exo-substituted fragments of 1a and 1b. It may also be expected that the bromo substituent on the B(2)position would also have some differential structural influences, and could contribute to variation of all the geometric parameters determined for 3b compared to those of unsubstituted 1a and 1b. In accord with this, for example, all of the interboron distances involving B(2) are rather shorter than the equivalent separations in 1a and 1b; in particular B(2)-B(6) is only 1.717(6) Å, short for such a connectivity.

It was previously reported that exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>arachno-B<sub>10</sub>H<sub>12</sub> 1b converts quantitatively into its exo,exo isomer 1a on heating in CD<sub>3</sub>C<sub>6</sub>D<sub>5</sub> solution. More exact work now shows that the reaction occurs with half-life  $t_1$  ca. 30 min in ca. 0.05 м CD<sub>3</sub>C<sub>6</sub>D<sub>5</sub> solution at 343 K. The 2-Br derivatives are marginally less labile, with  $t_1$  ca. 45 min at 343 K for the  $exo,endo \longrightarrow exo,exo$  conversion under the same conditions. By contrast, the dichloro species exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>-arachno-B<sub>10</sub>H<sub>10</sub> (compound 4b) seems much more stable, with no sign of change on heating at 343 K over similar periods to those that effect the quantitative isomerization of the monobromo compounds 3b and 3c. More extended heating appears to result in a more general decomposition. It may be noted that without rigorous exclusion of moisture these thermolyses result in boron-vertex loss to generate 4-(PMe<sub>2</sub>Ph)-arachno-B<sub>9</sub>H<sub>13</sub>. As with the initial exo, endo synthesis, the exo, endo  $\longrightarrow$  exo, exo conversions are quantitative and isomerically pure, suggesting no irreversible cluster-framework rearrangement during the conversion. The 6,9-L<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> system is quite robust when L is a ligand such as phosphine, so a dissociative mechanism for the  $exo,endo \longrightarrow exo,exo$  conversion is perhaps unlikely. Extrusion of the {BH(PMe<sub>2</sub>Ph)} vertex, rotation, and reincorporation is therefore a possibility. Here it may be pertinent that substantiated phosphine-induced vertex-extrusion mechanisms are recently precedented in boron-containing cluster chemistry.13

The NMR data for the exo, exo form of 6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-



**Fig. 4** Stick diagrams of the relative intensities and chemical shifts in the  $^{11}B$  NMR spectra of (from bottom to top)  $exo, exo-6, 9-(PMe_2Ph)_2-arachno-B_{10}H_{12}$  **1a**,  $exo-6, endo-9-(PMe_2Ph)_2-2-Br-arachno-B_{10}H_{11}$  **3b**,  $exo, exo-6, 9-(PMe_2Ph)_2-2-Br-arachno-B_{10}H_{11}$  **3a** and  $endo-6, exo-9-(PMe_2Ph)_2-2-Br-arachno-B_{10}H_{11}$  **3c**. Hatched lines join selected equivalent positions among the four species

arachno-B<sub>10</sub>H<sub>11</sub> 3a are given in Table 3 along with corresponding data for its endo, exo isomers 3b and 3c; NMR data for exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>-arachno-B<sub>10</sub>H<sub>10</sub> **4b** are given in Table 4. The <sup>11</sup>B spectra of the three 2-Br isomers 3a–3c are compared graphically with the spectrum<sup>8</sup> of unsubstituted exo,exo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> 1a in Fig. 4. Compared to its unsubstituted parent 1a there is a downfield shift of ca. 4 ppm for the bromine-substituted 2 position in 3a. This is within established ranges. 12,14 For the two exo,endo compounds 3b and 3c the resonances of the boron atoms B(2) and B(4) are shifted to low field relative to those of the exo, exo compound 3a. The magnitude of the downfield shift is 10.7 and 8.8 ppm for the boron atoms adjacent to the site of endo substitution [respectively B(4) in 3b and B(2) in 3c], and 2.9 and 3.6 ppm for those adjacent to the site of exo substitution [respectively B(2) in 3b and B(4) in 3c]. Similarly, atoms B(6) and B(9) bound to phosphorus are deshielded in 3b and 3c compared to 3a. If the phosphine is in the *endo* position this shift of the resonances to higher frequency is greater [4.0 ppm for B(9) in 3b and 5.9 ppm for B(6) in 3c] than if the phosphine is in the exo position [essentially unchanged at 0.5 ppm for B(6) in 3b and 0.6 ppm for B(9) in 3c]. A similar pattern of the chemical shifts would presumably occur in the dichlorodecaborane system, but in the absence of exo-exo-6-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>-arachno-B<sub>10</sub>H<sub>10</sub> 4a we are unable to confirm this.

The <sup>1</sup>H NMR properties of the {BH(PMe<sub>2</sub>Ph)} groupings at the 6 and 9 positions merit comment since these are instrumental in distinguishing the stereochemistry of the mixed species of the types 3a-3c in the absence of diffraction results. Comparison of the exo,endo forms of the parent species 6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> 1b with its symmetrical exo,exo analogue 1a readily shows that the {BH(exo)} unit of the endo-phosphinated 6 or 9 position resonates at  $\delta$ (<sup>1</sup>H) + 1.01, whereas the {BH(endo)} unit of an exo-phosphinated 6 or 9 position resonates at  $\delta$ (<sup>1</sup>H) - 1.41. This differential behaviour is in

accord with the general shielding behaviour of *endo versus exo* {BH} protons. <sup>14</sup> This behaviour, allied with the results of <sup>1</sup>H-{<sup>11</sup>B} correlation work, and of [<sup>1</sup>H-<sup>1</sup>H] and [<sup>11</sup>B-<sup>11</sup>B] correlation spectroscopy (COSY), is consistent with the assignment of the spectra and the consequent NMR substantiation of the three isomers **3a**–**3c** in the 6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-*arachno*-B<sub>10</sub>H<sub>11</sub> system (Table 3).

## **Experimental**

#### General

Reactions were carried out in dry solvents under dry nitrogen but subsequent manipulatory and separatory procedures were carried out in air. Preparative thin-layer chromatography (TLC) was carried out using 1.00 mm layers of silica gel G (Merck, type GF $_{254}$ ) made from water slurries on glass plates of dimensions  $20\times20~\text{cm}^2$ , followed by drying in air at 80 °C. The compound PMe $_2$ Ph was obtained commercially, and 2-BrB $_{10}H_{13}$  and 2,4-Cl $_2B_{10}H_{12}$  were prepared essentially as reported elsewhere.  $^{15}$ 

### Nuclear magnetic resonance spectroscopy

NMR spectroscopy was performed at ca. 5.9 and 9.4 T (fields corresponding to 250 and 400 MHz  $^{1}$ H frequencies respectively) using commercially available instrumentation and using techniques and procedures described and enunciated elsewhere. Shallow Chemical shifts  $\delta$  are given in ppm relative to  $\Xi = 100$  MHz for  $\delta(^{1}\text{H})$  ( $\pm$  0.05 ppm) (nominally SiMe<sub>4</sub>), 32.083 972 MHz for  $\delta(^{11}\text{B})$  ( $\pm$ 0.5 ppm) (nominally Et<sub>2</sub>O·BF<sub>3</sub> in CDCl<sub>3</sub>),  $^{14}$  and 40.480 730 MHz for  $\delta(^{31}\text{P})$  ( $\pm$  0.05 ppm) (nominally 85% aqueous H<sub>3</sub>PO<sub>4</sub>);  $\Xi$  is as defined in ref. 19.

#### **Preparations**

*exo,exo*-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> 1a and *exo,endo*-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>-arachno-B<sub>10</sub>H<sub>12</sub> 1b. A sample of B<sub>10</sub>H<sub>14</sub> (100 mg, 0.82 mmol) was dissolved in tetrahydrofuran (thf) (20 cm³), cooled to *ca.* 200 K, and then PMe<sub>2</sub>Ph (226 mg, 233 μl, 1.64 mmol) was added. The solution was stirred for 15 min at low temperature, then at room temperature for 2 h. The more volatile components were removed, the solid residue was redissolved in CH<sub>2</sub>Cl<sub>2</sub> (*ca.* 5 cm³) and the products were separated and purified by repeated preparative TLC, ultimate development with hexane–CH<sub>2</sub>Cl<sub>2</sub> (10:90) giving two products. These were exo,exo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>B<sub>10</sub>H<sub>12</sub> 1a (185 mg, 0.47 mmol, 57%;  $R_f$  0.85) and exo,endo-6,9-(PMe<sub>2</sub>Ph)<sub>2</sub>B<sub>10</sub>H<sub>12</sub> 1b (120 mg, 0.28 mmol, 34%;  $R_f$  0.70).

*exo*-6,*endo*-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-*arachno*-B<sub>10</sub>H<sub>11</sub> 3b and *endo*-6, *exo*-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-*arachno*-B<sub>10</sub>H<sub>11</sub> 3c. A sample of 2-BrB<sub>10</sub>H<sub>13</sub> (51 mg, 250 μmol) was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (20 cm<sup>3</sup>) at *ca.* 200 K, and then PMe<sub>2</sub>Ph (70 mg, 72 μl, 500 μmol) was added. The solution was stirred for 15 min at low temperature, allowed to warm to room temperature, and then stirred for 2 h. The more volatile components were removed, the solid residue was redissolved in 100% CH<sub>2</sub>Cl<sub>2</sub> (*ca.* 5 cm<sup>3</sup>) and the products were separated and purified by repeated preparative TLC, development with CH<sub>2</sub>Cl<sub>2</sub> giving two products. These were *exo*-6,*endo*-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-*arachno*-B<sub>10</sub>H<sub>11</sub> 3b (54 mg, 114 μmol, 45%;  $R_{\rm f}$  0.70) and *endo*-6,*exo*-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2-Br-*arachno*-B<sub>10</sub>H<sub>11</sub> 3c (52 mg, 110 μmol, 43%;  $R_{\rm f}$  0.56).

*exo*-6,*endo*-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>-*arachno*-B<sub>10</sub>H<sub>10</sub> 4b. A sample of 2,4-Cl<sub>2</sub>-*nido*-B<sub>10</sub>H<sub>12</sub> (36 mg, 190 μmol) was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (10 cm<sup>3</sup>), and then PMe<sub>2</sub>Ph (52 mg, 54 μl, 380 μmol) was added at *ca.* 200 K. The solution was stirred for 15 min at low temperature, allowed to warm to room temperature, and then stirred at room temperature for 2 h. Monitoring by <sup>11</sup>B NMR spectroscopy showed a quantitative conversion into the

Table 6 Crystallographic data and details of data collection and structure refinement for compounds 1a, 1b and 3b

	1a a	1b	3b
Formula	$C_{16}H_{34}B_{10}P_2$	$C_{16}H_{34}B_{10}P_2$	$C_{16}H_{33}B_{10}BrP_{2}$
$M_{\rm r}$	396.47	396.47	475.37
Crystal dimensions/mm	$0.89 \times 0.54 \times 0.12$	$0.79 \times 0.67 \times 0.53$	$0.54 \times 0.49 \times 0.42$
Crystal appearance	Colourless rod	Colourless block	Colourless block
Crystal system	Monoclinic	Triclinic	Monoclinic
Space group	$P2_1$	$P\bar{1}$	$P2_1/n$
a/Å	13.4155(9)	8.6670(4)	14.3789(12)
b/Å	12.0113(7)	10.3130(6)	11.8018(7)
c/Å	16.0081(12)	14.5912(9)	16.4193(12)
α/°	_	102.741(5)	_
β/°	113.704(6)	101.783(5)	114.418(7)
γ/°	_	103.293(5)	_
U/ų	2361.9(3)	1192.53(12)	2537.1(3)
Z	4	2	4
F(000)	840	420	976
$D_{\rm c}/{\rm Mg~m^{-3}}$	1.115	1.104	1.245
μ/mm <sup>-1</sup>	1.632	0.183	3.379
$\theta_{ m orientation}/^{\circ}$	30.1-34.9	15.5–17.5	33.1-41.7
θ <sub>data collection</sub> /°	3.01-64.34	2.11-24.97	3.45-64.41
h,k,l ranges	-15 to 15, $-13$ to 11, $-18$ to 18	-10 to 10, $-12$ to 12, $-17$ to 17	-16 to 16, $-13$ to 13, $-19$ to 19
Data measured	7803	8380	5308
Unique data, n	7227	4190	3892
Crystal decay (%)	9.2	0	3.7
$R_{\text{int}}^{\ b}$	0.0227	0.0199	0.0455
$T_{\min}$ , $T_{\max}$	0.723, 0.862	_	0.181, 0.358
Observed data $[F_0 > 2\sigma(I)]$	6962	3998	3779
$R1^c_{,} wR2^d_{,}$ (observed data)	0.0388, 0.1012	0.0302, 0.0829	0.0546, 0.1587
(all data)	0.0416, 0.1100	0.0333, 0.0888	0.0555, 0.1604
Goodness of fit on $F^2$ , $S^e$	1.062	1.002	1.150
Weighting parameters, $g_1, g_2^f$	0.0665, 0.9854	0.0457, 0.5289	0.0923, 4.1653
Number of parameters, p	609	305	310
Largest difference peak and hole/ e Å <sup>-3</sup>	0.27, -0.39	0.26, -0.32	0.80, -1.16

 $<sup>{}^</sup>a \text{ Flack absolute structure parameter} {}^{20} \text{ 0.01(2)}. \quad {}^b R_{\text{int}} = \Sigma |F_o^2 - F_o^2(\text{mean})|/\Sigma F_o^2. \quad {}^c R1 = \Sigma |F_o| - |F_c||/\Sigma |F_o|. \quad {}^d wR2 = [\Sigma w(F_o^2 - F_c^2)^2/\Sigma (F_o^2)^2]^{\frac{1}{2}}. \\ {}^c S = [\Sigma w(F_o^2 - F_c^2)^2/(n-p)]^{\frac{1}{2}}. \quad {}^f \text{ Weighting scheme } w^{-1} = [\sigma^2(F_o^2) + (g_1P)^2 + g_2P] \text{ where } P = (F_o^2 + 2F_c^2)/3.$ 

species isolated and identified as compound 4b. The more volatile components were removed, the solid residue was redissolved in CH<sub>2</sub>Cl<sub>2</sub> (ca. 5 cm<sup>3</sup>) and subjected to preparative TLC, development with CH2Cl2 giving one product only. This was exo-6,endo-9-(PMe<sub>2</sub>Ph)<sub>2</sub>-2,4-Cl<sub>2</sub>-arachno-B<sub>10</sub>H<sub>10</sub> **4b** (81 mg, 174  $\mu$ mol, 93%;  $R_{\rm f}$  0.75).

### Crystallography

Crystals of compounds 1a, 1b and 3b suitable for single-crystal X-ray diffraction analysis were each grown from CH<sub>2</sub>Cl<sub>2</sub>hexane at room temperature. All crystallographic measurements were carried out at 160(2) K on a Stoe STADI 4 fourcircle diffractometer operating in the  $\omega$ - $\theta$  scan mode using graphite-monochromated X-radiation (Cu-K $\alpha$ ,  $\overline{\lambda}$  = 1.541 86 Å for **1a** and **3b**; Mo-K $\alpha$ ,  $\overline{\lambda} = 0.710 73 \text{ Å for$ **1b** $}). Crystal data and$ refinement parameters for the three crystals are listed in Table 6. Cell dimensions were refined from the values of 50 (for 1a), 70 (for 1b) or 30 (for 3b) selected reflections (together with their Friedel opposites) that were well separated in reciprocal space and that were measured at  $\pm \theta$  in order to minimize systematic errors. The three sets of data were corrected for Lorentz and polarization effects. An empirical absorption correction based on  $\psi$  scans was applied to the data sets collected for compounds 1a and 3b. The structures of the three compounds were solved by direct methods using SHELXS 86<sup>21</sup> and were refined by fullmatrix least squares (against all the unique  $F^2$  data) using SHELXL 93.22 Refinement was similar for all three structures. Two independent molecules of compound 1a are present in the asymmetric unit. All non-hydrogen atoms were refined with anisotropic displacement parameters. Phosphine hydrogen atoms were constrained to idealized positions with a riding model that included free rotation of methyl groups and with PPh units restrained to be flat and of overall  $C_{2v}$  symmetry.

Cluster hydrogen atoms were located via Fourier-difference syntheses and freely refined with isotropic displacement parameters. The crystallographically imposed absolute structure of the crystal of compound 1a was confirmed by the refinement of the Flack absolute structure parameter  $^{20}$  to 0.01(2).

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